Analgesic Serotonin from Banana Fruit (*Musa paradisiaca*) on Serotonin 1 b (5-HT1b) Receptor Protein *In Silico*

Rahadian Zainul^{a,b,*}^(b)| Herland Satriawan^(b)| Dheo Shalsabilla Novel^{a,b}^(b)| Rismi Verawati^{a,b} ^(b)| Amalia Putri Lubis^{a,b}^(b)| Vikash Jakhmola^d^(b)| Meksim Rebezov^{e,f}^(b)| Syafrizal Syafrizal^g^(b)| Shafique Ahmed^b^(b)| Mishra Lakshmi^(b)

^aDepartment of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Negeri Padang, Padang, West Sumatra, Indonesia ^bCenter for Advanced Material Processing, Artificial Intelligence and Biophysics Informatics (CAMPBIOTICS), Universitas Negeri Padang, Indonesia

^cInstitute of Ocean and Earth Sciences, University of Malaya, 50603, Kuala Lumpur, Malaysia

^dUttaranchal Institute of Pharmaceutical Sciences, Uttaranchal University, Dehradun, India

^eDepartment of Scientific Research, V. M. Gorbatov Federal Research Center for Food Systems, 26 Talalikhin Str., Moscow 109316, Russia ^fFaculty of Biotechnology and Food Engineering, Ural State Agrarian University, 42 Karl Liebknecht str., Yekaterinburg, 620075, Russia ^gDepartment of Animal Husbandry, Faculty of Agricultural, Universitas Tamansiswa Padang, Indonesia

^hDepartment of Biological Sciences, Faculty of Allied Health Sciences, Faculty of Allied Health Sciences, The Superior University, Lahore, Pakistan

International School of Asfendiyarov Kazakh National Medical University, Almaty, Kazakhstan



<u>Citation</u> R. Zainul,H. Satriawan, D. Shalsabilla Novel, R. Verawati, A. Putri Lubis, V. Jakhmola, M. Rebezov, Syafrizal **Analgesic Serotonin from Banana Fruit (***Musa paradisiaca***) on Serotonin 1 b (5-HT1b) Receptor Protein** *In Silico. J. Appl. Organomet. Chem.***, 2024**, 4(2), 88-99.

400 https://doi.org/10.48309/JAOC.2024.443746.1166



Article info: Submitted: 1 February 2024 Revised: 21 February 2024 Accepted: 05 March 2024

ID: JAOC-2402-1166 Checked for Plagiarism: Yes Language Editor Checked: Yes

Keywords:

Serotonin analgesic, Banana fruit (Musa paradisiaca), Serotonin 1 b (5-HT1b) receptor protein, Molecular interaction, *In silico* approach

<u>A B S T R A C T</u>

Serotonin analgesics from banana (Musa paradisiaca) fruit have been investigated to determine potential interactions with serotonin 1 b (5-HT1b) receptors at the molecular level. The study utilized an *in silico* approach to predict the interaction between serotonin analgesics and receptor proteins. The research method involved the use of Pymol, MOE 2015, Discovery Studio, and Lipinski Rule software. The use of Pymol and MOE was used for visualization of the molecular structures of serotonin analgesics and receptor proteins. Discovery Studio was used to analyze the interaction between serotonin analgesic and receptor protein, which revealed the presence of binding between the two with Binding Affinity of -5.1297 and -11.1061 and RMSD of 1.7373 and 3.7057. In addition, analysis by Lipinski Rule revealed the molecular characteristics of the serotonin analgesic, including a mass of 196, no hydrogen bond donor, two hydrogen bond acceptors, a log P of 3.023, and a molar reactivity of 56.390. These results demonstrate the analgesic potential of serotonin in interacting with serotonin 1 b (5-HT1b) receptors, which may form the basis for further research in drug development related to serotonin-based pain treatment.

Introduction

B ananas (*Musa paradisiaca*) have long been one of the most popular and widely consumed fruits around the world. In addition to its great taste and nutrient-rich content, bananas have also been recognized for their potential health properties. One of the components of interest in bananas is the analgesic serotonin [1-3]. Serotonin is a neurotransmitter that plays an important role in regulating various neurobiological and physiological functions in humans. One of the known types of serotonin receptors is the 5-HT1b receptor. This receptor is mainly found in the brain and is a target for the treatment of various neurological and psychiatric disorders, including pain and depression [4, 5].

Pain is one of the most common health problems experienced bv humans. The development of effective and safe analgesics is a major focus in pharmaceutical research. One promising approach is through the interaction between serotonin analgesics and serotonin 1 b (5-HT1b) receptor proteins [6, 7]. Banana fruit (Musa paradisiaca) is known to contain serotonin compounds that have potential as analgesics. Therefore, this study aims to understand the molecular interaction between serotonin analgesics derived from banana fruit and serotonin receptor protein 1 b (5-HT1b) in silico [8-10].

Research into serotonin analgesics and interactions with serotonin 1 b (5-HT1b) receptors is still in its infancy. Some previous studies have shown that serotonin compounds have potential as analgesics, and banana fruit (Musa paradisiaca) is known to contain significant amounts of serotonin [11-13]. In *silico* approaches using software such as Pymol, Molecular Operation Environment 2015.10 (MOE 2015), Discovery Studio 2016, and Lipinski Rule can provide insights into the molecular interactions between serotonin analgesics and receptor proteins, which could pave the way for new drug development in serotonin-based pain treatment [14-16].

The novelty of this study is the use of an *in silico* approach to study the interaction between banana-derived serotonin analgesics and serotonin receptor protein 1 b (5-HT1b). In the context of analgesic development, previous studies have identified the analgesic potential of serotonin and serotonin content in banana fruit [17-18]. However, this study combines an *in silico* approach with the use of software such as Pymol, MOE 2015, Discovery Studio 2016 and Lipinski Rule to analyze molecular interactions in depth [19-21].

The contribution of this study is to provide a more comprehensive understanding of the analgesic potential of banana-derived serotonin and its molecular interactions with serotonin receptor protein 1 b (5-HT1b), which can serve as a foundation for the development of more effective and safe serotonin-based pain medications [22, 23]. This study aims to understand the molecular interactions between serotonin analgesics derived from banana fruit and serotonin receptor protein 1 b (5-HT1b) *in silico* to identify their analgesic potential and provide new insights in the development of serotonin-based pain medications.

Experimental

Materials and Methods

The research method used in this study consists of several stages. The initial stage is molecular data collection. The molecular structure of serotonin analgesic from banana (Musa paradisiaca) is obtained from reliable literature sources or molecular synthesis can be done using software such as ChemDraw (https://www.cambridgesoft.com/Ensemble_fo r_Chemistry/ChemDraw/). Data on serotonin receptor protein 1 b (5-HT1b) can be obtained from protein databases such as Protein Data Bank (PDB) (https://www.rcsb.org/) using the appropriate PDB code [24-25].

The next step is visualization and analysis of the molecular structure. Pymol (https://pymol.org/) and MOE 2015 software can be used to visualize the molecular structures of serotonin analgesics and serotonin 1 b (5-HT1b) receptor proteins. This allows researchers to understand important conformations and structural features [26, 27].

Afterwards, molecular interaction analysis was performed using Discovery Studio 2016. This software can facilitate the modeling of the interaction between serotonin analgesics and receptor proteins. The results of the analysis will provide information regarding Binding Affinity and RMSD, which indicate the strength of the bond and the stability of the interaction between the two molecules [28-32].

The last stage is Lipinski rule analysis. Lipinski rule analysis is performed to examine the molecular characteristics of serotonin analgesics. Lipinski Rule software can be used to analyze factors such as molecular mass, hydrogen bond donor, hydrogen bond acceptor, log P, and molar reactivity. Using this method, researchers can gain a

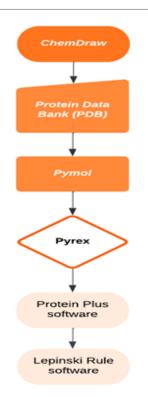


Figure 1. Flowchart research

deeper understanding of the molecular interactions between serotonin analgesics from banana fruit and serotonin 1 b (5-HT1b) receptor proteins, contributing to the interpretation of data and the development of more effective serotonin-based pain medications [33-35].

Results and Discussion

The results of this study involve several important aspects related to the molecular interaction between serotonin analgesics from banana fruit and serotonin receptor protein 1 b (5-HT1b). Molecular interaction analysis using Discover Studio 2016, this study successfully analyzed the interaction between serotonin analgesic and serotonin 1 b (5-HT1b) receptor protein. The results of this analysis revealed a

binding between serotonin analgesics and serotonin receptors, which was reinforced by the Binding Affinity values found (-5.1297 and -11.1061). The docking conformation of dihydroergotamine has a hydrogen bond with Val201. On the other hand, no hydrogen bond was detected from Xanthone. However, due to the high value of RMSD (< 2 Å) on dihydroergotamine shows a poor dock conformation result [36].

This suggests that serotonin analgesics have the potential to interact with 5-HT1b receptors, which are important targets in analgesic drug development [37-39]. Table 1 indicates the molecular docking analysis result of Xanthone and ligand reference on 5-HT1b receptor protein. Analyzing molecular characteristics through Lipinski rule analysis using Lipinski Rule software, this study examined the molecular characteristics of serotonin analgesic. In this analysis, it was found that serotonin analgesic has a mass of 196, no hydrogen bond donor, two hydrogen bond acceptors, a log P of 3.023, and a molar reactivity of 56.390. These results provide a deeper understanding of the chemical properties and molecular characteristics of serotonin analgesics, which can be an important basis in the development of effective analgesic drugs [40, 41]. Molecular visualization analysis with the use of Pymol and MOE 2015 software allows visualization of the molecular structure of serotonin analgesics and serotonin 1 b (5-HT1b) receptor proteins. With the help of these visualizations, researchers can understand the conformation and relevant structural features. This visual analysis helps in understanding the interaction between the two molecules and gives a clearer picture of how serotonin analgesics can potentially bind to the 5-HT1b receptor [42, 43]. Table 2 presents the data from Lipinski and Figure 1 displays the interaction results of Serotonin and 4iaq.

Ligand	Binding Affinity (Kcal/mol)	RMSD (Å)	Hydrogen bond	Van der Waals interaction	Carbon hydrogen bond	Amino acid residue (hydrogen bond)
Dihydroergota mine (Ligand reference)	-11.1061	3.7057	1	16	1	Val201
Xanthone	-5.1297	1.7373	0	9	0	-
Not available						

Table 1. Molecular docking analysis result

Journal of Applied Organometallic Chemistry

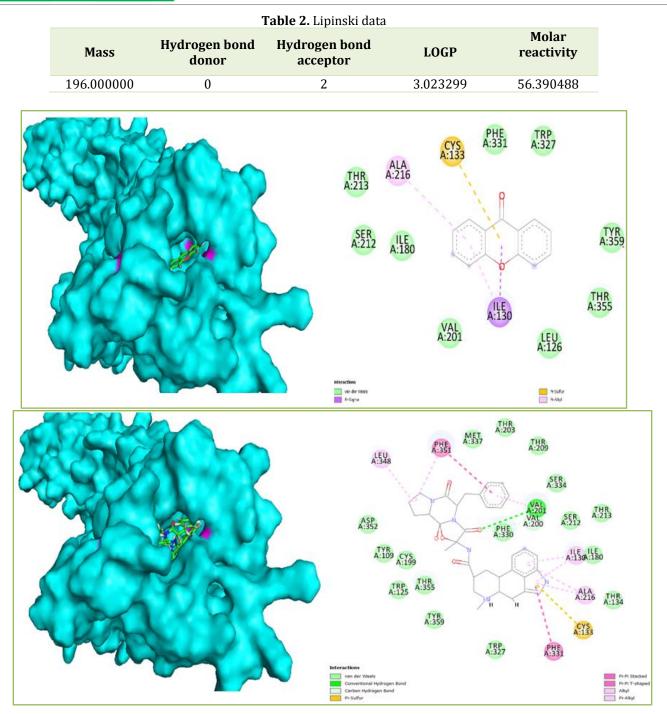


Figure 2. Xanthone (above) and dihydroergotamine (below) visual and interactions

Overall, this analysis provides valuable insights into the molecular interactions between serotonin analgesics from banana fruit with serotonin receptor protein 1 b (5-HT1b). These findings may make important contributions to the development of new serotonin-based analgesic drugs, with a better understanding of the molecular characteristics, interactions, and therapeutic potential of serotonin analgesics in pain relief [4-7].

The interpretation of this research led to an important understanding of the analgesic potential of banana (*Musa paradisiaca*) fruit-derived serotonin in binding to the serotonin 1 b (5-HT1b) receptor protein [2,6,10]. Using *in silico* approaches and software such as Pymol,

MOE 2015, Discovery Studio, and Lipinski Rule, this study revealed the presence of interactions between serotonin analgesics and receptor proteins, with Binding Affinity indicating significant binding strength. These results suggest that serotonin analgesics from banana fruit have the potential to function as painrelieving agents through interaction with the 5-HT1b receptor [44, 45].

In addition, analysis of the molecular characteristics of serotonin analgesics showed that these compounds have physicochemical properties that conform to the requirements of the Lipinski rule, such as adequate molecular mass, optimal log P, and the presence of hydrogen bond acceptors that facilitate interaction with receptors. This suggests that serotonin analgesics from banana fruit have favorable pharmacokinetic potential to be developed as analgesic drugs [46, 47].

Overall, the interpretation of this research suggests that banana fruit has potential as a natural source of serotonin analgesics that can interact with serotonin 1 b (5-HT1b) receptors. discoverv provides This an important foundation for continued research in the development of more effective and safe serotonin-based drugs in the treatment of pain. With a better understanding of the molecular interactions and molecular characteristics of serotonin analgesics, this research provides a valuable contribution in the search for better treatment solutions to address the pain problems that humans often face [48, 49].

Comparisons of this research from several perspectives and reviews can provide a deeper understanding of the contributions and uniqueness of this research. In terms of research methods, this research utilizes in-silico approaches and software such as Pymol, MOE 2015, Discovery Studio 2016, and Lipinski Rule. This approach enables efficient analysis of molecular interactions molecular and characteristics of serotonin analgesics. Compared to *in vitro* or *in vivo* experimental studies, in silico approaches have advantages in terms of cost and time, as well as allowing efficient preliminary testing before conducting further experiments. However, it is also important to remember that in silico findings

need to be verified with actual experiments to validate the results [50-51].

This study shows the potential application of serotonin analgesics from banana fruit in the development of serotonin-based pain medications. These findings can serve as a foundation for the development of new analgesic drugs that are potentially more effective and safe. The utilization of natural sources such as banana fruit as a basic ingredient for analgesics can also be an attractive alternative in the pharmaceutical industry. However, further research is needed to understand the efficacy, bioavailability, and safety of serotonin analgesics from banana fruit before they can be used clinically [2,8,10].

In scientific terms, this study contributes to broadening the understanding of the molecular interactions and molecular characteristics of serotonin analgesics. Through the analysis of molecular interactions and characteristics, this study provides important insights into the uniqueness potential and of serotonin analgesics from banana fruit. These findings may encourage further research in the field of pain medicine development and enrich the scientific literature related to molecular interactions between serotonin analgesics and serotonin 1 b (5-HT1b) receptors (Figure 2) [6, 7,44].

Overall, this research has comparative value in research methods, terms of potential applications, and scientific contributions. In the context of serotonin-based analgesic drug development, this research provides an important contribution in understanding the molecular interactions and molecular characteristics of serotonin analgesics from banana fruit. However, further research and experimental validation are needed to strengthen these findings and encourage practical application in pain treatment [52, 53]. Figure 3 and Figure 4 demonstrate the Xanthone ligand and 4iaq protein.

The recent research on serotonin receptor 1B (5-HT1B) has provided significant insights into its structure and function, which can be compared with findings from previous studies. The study by Xu *et al.*, published in Nature, has revealed crucial details about the molecular underpinnings of 5-HT receptor activation and

Journal of Applied Organometallic Chemistry

modulation. This studv highlights the importance of cholesterol in the function and organization of G Protein-Coupled Receptors (GPCRs), including the 5-HT1B receptor. Cholesterol was found to make direct contact with the receptor and contribute to the high affinity of ligands, indicating its vital role in receptor activity. In addition, the study discussed the importance of palmitoylation at the receptor's C-terminal cysteine residues for pointing towards efficient signaling, the synergistic effects of cholesterol and palmitoylation on receptor structure and function [54].

Another study focused on the crystal structure of the human 5-HT1B serotonin receptor bound to an inverse agonist, providing detailed insights into the receptor's structural aspects that facilitate drug binding and receptor modulation. The use of the fusion partner OB1 to improve crystallization was highlighted, showcasing the technical advancements in understanding receptor-ligand interactions at the molecular level. These findings underline the complex interplay of structural modifications and their impact on the receptor's pharmacological properties [54].

Comparing these findings with previous research underscores the evolution of our understanding of 5-HT1B receptors. The detailed structural insights from recent studies, such as those on cholesterol's role and the effect of palmitoylation and crystallization techniques, offer a more nuanced understanding of receptor function and its implications for drug development and therapeutic interventions. This represents a significant advancement over earlier studies that might not have had the benefit of such detailed molecular and structural insights [54].

The recent advancements in research on the receptor 1b (5-HT1b) have serotonin highlighted the intricate mechanisms of receptor activation and the structural basis for ligand recognition and receptor modulation. A study detailed the structural insights into the lipid and ligand regulation of serotonin receptors, focusing on the functional importance of specific interactions between lipids and G protein-coupled receptors (GPCRs). This research emphasized the role of membrane-lipid interactions in regulating GPCR dynamics and functions, with particular attention to how phospholipids like phosphatidylinositol 4-phosphate (PtdIns4P) enhance the activity of the receptor-G protein complex, acting as positive allosteric modulators. The study further explored the contribution of cholesterol to receptor activity, highlighting the need for additional experiments to understand its role in the structure of the ligand-binding pocket of 5-HT receptors (Signal Transduction and Targeted Therapy) [55,56].

Another study focused on the engineering of a BRIL fusion partner to facilitate the crystallization of the 5-HT1B receptor in complex with an inverse agonist. This research aimed to improve the diffraction quality of the crystals by optimizing the sequence of BRIL, introducing mutations to reduce surface entropy and enhance specific polar interactions. The study successfully obtained a crystal structure that provided insights into the receptor's ligand-binding affinity and the effects specific mutations on crystallization of efficiency (Cell Discovery) [54].

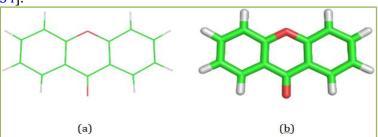


Figure 3. (a) 2D Visualization of xanthone ligand (b) 3D visualization of xanthone ligand

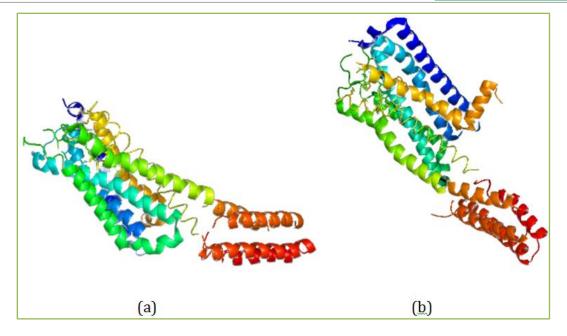


Figure 4. (a) Net protein serotonin 1b (5-HT1b) receptor (b) P net protein serotonin 1b (5-HT1b) receptor

Furthermore, cryo-electron microscopy (cryo-EM) has been employed to elucidate the structure of the 5-HT1B receptor coupled to heterotrimeric Go, offering a detailed view of the receptor's architecture and its interaction with G proteins. This approach has opened new avenues for understanding the dynamic processes involved in receptor activation and signal transduction, highlighting the receptor's conformational changes upon ligand binding and G protein coupling (Nature) [57].

Conclusion

The conclusion of this study is that serotonin analgesics derived from banana fruit (Musa paradisiaca) show potential in interacting with serotonin 1 b (5-HT1b) receptor protein in provides a silico. This study deeper understanding of the molecular interactions and characteristics molecular of serotonin analgesics, as well as their potential application in the development of serotonin-based analgesic drugs. These findings make an important contribution to pain research and drug development, but further research and experimental validation are needed to verify the results and understand the therapeutic potential and safety of serotonin analgesics

from banana fruit before they can be applied clinically.

Acknowledgements

The authors would like to express their profound gratitude to Universitas Negeri Padang for its invaluable support and contributions to this research. The guidance, resources, and facilities provided by the university have been instrumental in the completion of this study.

Disclosure Statement

The authors have declared that there is no competing interest in this article

Orcid

Rahadian Zainul *https://orcid.org/0000-0002-3740-3597*

Herland Satriawan https://orcid.org/0009-0007-2398-4793

Dheo Shalsabilla Novel https://orcid.org/0009-0006-0145-5180

Rismi Verawati https://orcid.org/0009-0006-2143-0300

Journal of Applied Organometallic Chemistry

Amalia Putri Lubis https://orcid.org/0009-0002-7797-2492

Vikash Jakhmola https://orcid.org/0000-0002-8108-006X

Meksim Rebezov https://orcid.org/0000-0003-0857-5143

Syafrizal Syafrizal https://orcid.org/0009-0004-7764-9956

Shafique Ahmed *https://orcid.org/0009-0001-2463-6316*

Mishra Lakshmi https://orcid.org/0009-0007-4042-9580

References

[1]. V. Galani, Musa paradisiaca Linn.-A comprehensive review, *Scholars International Journal of Traditional and Complementary Medicine*, **2019**, 45-56. [Crossref], [Google Scholar]

[2]. A. Mohiuddin, M.K. Saha, M.S. Hossian, A. Ferdoushi, Usefulness of banana (Musa paradisiaca) wastes in manufacturing of bioproducts: A review, *Agriculturists*, **2014**, *12*, 148-158. [Crossref], [Google Scholar]

[3]. N. Rajesh, Medicinal benefits of Musa paradisiaca (Banana), *International Journal of Biology Research*, **2017**, *2*, 51-54. [Crossref], [Google Scholar]

[4]. J. García-Nafría, R. Nehmé, P.C. Edwards, C.G. Tate, Cryo-EM structure of the serotonin 5-HT1B receptor coupled to heterotrimeric G_o, *Nature*, **2018**, *558*, 620-623. [Crossref], [Google Scholar], [Publisher]

[5]. D. David, A. Gardier, The pharmacological basis of the serotonin system: Application to antidepressant response, *L'encephale*, **2016**, *42*, 255-263. [Crossref], [Google Scholar], [Publisher]

[6]. E. Silk, M. Diwan, T. Rabelo, H. Katzman, A.C.P. Campos, F.V. Gouveia, P. Giacobbe, N. Lipsman, C. Hamani, Serotonin 5-HT1B receptors mediate the antidepressant-and anxiolytic-like effects of ventromedial prefrontal cortex deep brain stimulation in a mouse model of social defeat, *Psychopharmacology*, **2022**, *239*, 3875-3892. [Crossref], [Google Scholar], [Publisher]

[7]. L. Fang, S. He, P. Yin, N. Wang, B. Zhang, H. Jin, Design, synthesis, and structure–Activity relationship studies of novel tryptamine derivatives as 5-HT1B receptor agonists, *Journal of Molecular Structure*, **2022**, *1265*, 133320. [Crossref], [Google Scholar], [Publisher]

[8]. S. Sahebnasagh, J. Fadaee Kakhki, M. Ebrahimi, M.R. Bozorgmehr, M.R. Abedi, Preconcentration and determination of fluoxetine in hospital wastewater and human hair samples using solid-phase μ-extraction by silver nanoparticles followed by spectro-fluorimetric, *Chemical Methodologies*, **2021**, *5*, 211-218. [Crossref], [Google Scholar], [Publisher]

[9]. D. Kemisetti, D.B.B. Rajeswar Das, A comprehensive review on Musa Paradisiaca taxonomical, morphological classification and its pharmacological activities, *Journal of Pharmaceutical Negative Results*, **2022**, 737-749. [Crossref], [Google Scholar], [Publisher]

[10]. A. Rana, A. Goel, S. Yadav, S. Praveen, T. Kumar, A review on pharmacological activities of banana (Musa Paradisiaca), *International Journal of Pharmaceutical Research and Applications*, **2023**, *8*, 2901-2910. [Crossref], [Publisher]

[11]. S. Abbasbeigi, Misfolded structures|A brief insight into protein aggregation criteria, which may lead to Proteopathy diseases, *Journal of Chemical Reviews*, **2021**, *3*, 97-108. [Crossref], [Google Scholar], [Publisher]

[12]. B.E. Ehigiator, N.C. Offonry, E. Adikwu, B.O. Inemesit, Safety, anti-inflammatory and analgesic assessments of methanolic extract of Musa paradisiaca peel in Sprague Dawley rats, *African Journal of Pharmacology and Therapeutics*, **2018**, *7*. [Google Scholar], [Publisher]

[13]. H.S. Budi, W.S. Juliastuti, Y.P. Sulistyowati, The bleeding and clotting time analysis of the stem extract of Musa paradisiaca var. sapientum (L.) Kunze on hemostatic response, *Jordan Journal of Pharmaceutical Sciences*, **2021**, *14*, 1-8. [Google Scholar], [Publisher]

[14]. N.S. Aini, V.D. Kharisma, M.H. Widyananda, A.A.A. Murtadlo, R.T. Probojati, D.D.R. Turista, M.B. Tamam, V. Jakhmola, E. Yuniarti, S. Al Aziz, In silico screening of bioactive compounds from Garcinia mangostana L. against SARS-CoV-2 via tetra inhibitors, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[15]. A.T. Rahman, A. Jethro, P. Santoso, V.D. Kharisma, A.A.A. Murtadlo, D. Purnamasari, N.H. Soekamto, A. Ansori, R.S. Mandeli, K.A.M.S. Aledresi, In silico study of the potential of endemic sumatra wild turmeric rhizomes (Curcuma Sumatrana: Zingiberaceae) as anticancer, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[16]. N. Mawaddani, E. Sutiyanti, M.H. Widyananda, V.D. Kharisma, D.D.R. Turista, M.B. Tamam, V. Jakhmola, B.R. Fajri, M.R. Ghifari, M.T. Albari, In silico study of entry inhibitor from Moringa oleifera bioactive compounds against SARS-CoV-2 infection, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[17]. S.P. Bangar, N. Sharma, H. Kaur, M. Kaur, K.S. Sandhu, S. Maqsood, F. Ozogul, A review of sapodilla (Manilkara zapota) in human nutrition, health, and industrial applications, *Trends in Food Science & Technology*, **2022**, *127*, 319-334. [Crossref], [Google Scholar], [Publisher]

[18]. B. Salehi, F. Sharopov, P.V.T. Fokou, A. Kobylinska, L.d. Jonge, K. Tadio, J. Sharifi-Rad, M.M. Posmyk, M. Martorell, N. Martins, Melatonin in medicinal and food plants: Occurrence, bioavailability, and health potential for humans, *Cells*, **2019**, *8*, 681. [Crossref], [Google Scholar], [Publisher]

[19]. M.E. Ullah, R.T. Probojati, A.A.A. Murtadlo, M.B. Tamam, S.W. Naw, Revealing of antiinflamatory agent from zingiber officinale var. Roscoe via IKK-B inhibitor mechanism through in silico simulation, *SAINSTEK* International Journal on Applied Science, Advanced Technology and Informatics, **2022**, 1, 14-19. [Crossref], [Google Scholar], [Publisher]

[20]. P. Listiyani, V.D. Kharisma, A.N.M. Ansori, M.H. Widyananda, R.T. Probojati, A.A.A. Murtadlo, D.D.R. Turista, M.E. Ullah, V. Jakhmola, R. Zainul, In silico phytochemical compounds screening of Allium sativum targeting the Mpro of SARS-CoV-2, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[21]. P. Listiyani, S.L. Utami, D.D.R. Turista, A. Wiguna, A. Wijayanti, Y. Rachmawati, A.F. Dibha, T. Hasan, M.A. Hafidzhah, R.M. Wijaya, Computational screening of toxicity, drug-like molecule, and bioactivity from green tea phytochemical as antiviral candidate, *SAINSTEK International Journal on Applied Science, Advanced Technology and Informatics*, **2022**, *1*, 39-45. [Crossref], [Google Scholar]

[22]. W. Yin, X.E. Zhou, D. Yang, P.W. de Waal, M. Wang, A. Dai, X. Cai, C.-Y. Huang, P. Liu, X. Wang, Crystal structure of the human 5-HT1B serotonin receptor bound to an inverse agonist, *Cell discovery*, **2018**, *4*, 12. [Crossref], [Google Scholar], [Publisher]

[23]. C. Gadgaard, A.A. Jensen, Functional characterization of 5-HT1A and 5-HT1B serotonin receptor signaling through G-protein-activated inwardly rectifying K⁺ channels in a fluorescence-based membrane potential assay, *Biochemical pharmacology*, **2020**, *175*, 113870. [Crossref], [Google Scholar], [Publisher]

[24]. L. Van Huyen, T.Q. Toan, P.Q. Long, P.M. Quan, Computer aided screening of indirubin analogues targeting GSK-3β protein using molecular docking, *Biomedical Journal of Scientific & Technical Research*, **2019**, 14844-14845. [Crossref], [Google Scholar], [Publisher]

[25]. V.D. Kharisma, A.N.M. Ansori, F.A. Dian, W.C. Rizky, T.G.A. Dings, R. Zainul, A.P. Nugraha, Molecular docking and dynamic simulation of entry inhibitor from tamarindus indica bioactive compounds against SARS-CoV-2 infection via viroinformatics study, *Biochemical and Cellular Archives*, **2021**, *21*, 3323-3327. [Crossref], [Google Scholar] [26]. N.S. Aini, V.D. Kharisma, M.H. Widyananda, A.A.A. Murtadlo, R.T. Probojati, D.D.R. Turista, M.B. Tamam, V. Jakhmola, D.P. Sari, M.T. Albari, In silico screening of bioactive compounds from Syzygium cumini L. and moringa oleifera L. against SARS-CoV-2 via tetra inhibitors, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[27]. A.A. Rabaan, M.A. Halwani, M. Aljeldah, B.R. Al Shammari, M. Garout, J. Aldali, A. Alawfi, A. Alshengeti, A.M. Alsulaiman, A. Alsayyah, Exploration of potent antiviral phytomedicines from Lauraceae family plants against SARS-CoV-2 RNA-dependent RNA polymerase, *Journal of Biomolecular Structure and Dynamics*, **2023**, 1-21. [Crossref], [Google Scholar], [Publisher]

[28]. F. Maryam, H. Mukhtar, I. Bibi, M. Rizwan, S. Khan, A. Mehmood, Ligand based pharmacophore modelling, virtual screening and molecular docking of novel compounds against diabetes, *Bulletin (Eugenics Society (London, England))*, **2019**, *8*, 38-48. [Crossref], [Publisher]

[29]. M. Réau, F. Langenfeld, J.-F. Zagury, N. Lagarde, M. Montes, Decoys selection in benchmarking datasets: overview and perspectives, *Frontiers in Pharmacology*, **2018**, *9*, 11. [Crossref], [Google Scholar], [Publisher]

[30]. L. Pinzi, G. Rastelli, Molecular docking: shifting paradigms in drug discovery, *International Journal of Molecular Sciences*, **2019**, *20*, 4331. [Crossref], [Google Scholar], [Publisher]

[31]. R. Selvaraj, G. Hemalatha, K. Sivakumari, In silico molecular docking stuides of Muricin J, Muricin K and Muricin L compound from A. muricata against apoptotic proteins (caspase-3, caspase-9 and $\hat{1}^2$ -actin), *Innoriginal: International Journal of Sciences*, **2020**, 1-4. [Google Scholar], [Publisher]

[32]. X. Lin, X. Li, X. Lin, A review on applications of computational methods in drug screening and design, *Molecules*, **2020**, *25*, 1375. [Crossref], [Google Scholar], [Publisher]

[33]. H. Patel, A. Kukol, Integrating molecular modelling methods to advance influenza A virus drug discovery, *Drug Discovery Today*, **2021**, *26*, 503-510. [Crossref], [Google Scholar], [Publisher]

[34]. A. Dibha, S. Wahyuningsih, A. Ansori, V. Kharisma, M. Widyananda, A. Parikesit, M. Sibero, R. Probojati, A. Murtadlo, J. Trinugroho, Utilization of secondary metabolites in algae Kappaphycus alvarezii as a breast cancer drug with a computational method, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[35]. O.A. Ojo, A.B. Ojo, C. Okolie, M.-A.C. Nwakama, M. Ivobhebhe, I.O. Evbuomwan, C.O. Nwonuma, R.F. Maimako, A.E. Adegboyega, O.A. Taiwo, Deciphering the interactions of bioactive compounds in selected traditional medicinal against Alzheimer's diseases plants via pharmacophore modeling, auto-QSAR, and molecular docking approaches, Molecules, 2021, [Crossref]. [Google 26, 1996. Scholar]. [Publisher]

[36]. S.A. Attique, M. Hassan, M. Usman, R.M. Atif, S. Mahboob, K.A. Al-Ghanim, M. Bilal, M.Z. Nawaz, A molecular docking approach to evaluate the pharmacological properties of natural and synthetic treatment candidates for use against hypertension, *International Journal of Environmental Research and Public Health*, **2019**, *16*, 923. [Crossref], [Google Scholar], [Publisher]

[37]. M.B. de Ávila, W.F. de Azevedo Jr, Development of machine learning models to predict inhibition of 3-dehydroquinate dehydratase, *Chemical Biology & Drug Design*, **2018**, *92*, 1468-1474. [Crossref], [Google Scholar], [Publisher]

[38]. G. Xiong, Z. Wu, J. Yi, L. Fu, Z. Yang, C. Hsieh, M. Yin, X. Zeng, C. Wu, A. Lu, ADMETlab 2.0: an integrated online platform for accurate and comprehensive predictions of ADMET properties, *Nucleic Acids Research*, **2021**, *49*, W5-W14. [Crossref], [Google Scholar], [Publisher] [39]. J. Lemkul, From proteins to perturbed Hamiltonians: a suite of tutorials for the GROMACS-2018 molecular simulation package [Article v1. 0]. *Living Journal of Computational Molecular Science*, **2019**, *1*, 1–53. [Crossref], [Google Scholar], [Publisher]

[40]. J. Han, L. Geng, C. Lu, J. Zhou, Y. Li, T. Ming, Z. Zhang, X. Su, Analyzing the mechanism by which oyster peptides target IL-2 in melanoma cell apoptosis based on RNA-seq and m6A-seq, *Food & Function*, **2023**, *14*, 2362-2373. [Crossref], [Google Scholar], [Publisher]

[41]. A. Ansori, V. Kharisma, A. Parikesit, F. Dian, R. Probojati, M. Rebezov, P. Scherbakov, P. Burkov, G. Zhdanova, A. Mikhalev, Bioactive compounds from mangosteen (Garcinia mangostana L.) as an antiviral agent via dual inhibitor mechanism against SARSCoV-2: An in silico approach, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[42]. N.R. Jabir, M.T. Rehman, K. Alsolami, S. Shakil, T.A. Zughaibi, R.F. Alserihi, M.S. Khan, M.F. AlAjmi, S. Tabrez, Concatenation of molecular docking and molecular simulation of BACE-1, γ-secretase targeted ligands: in pursuit of Alzheimer's treatment, *Annals of Medicine*, **2021**, *53*, 2332-2344. [Crossref], [Google Scholar], [Publisher]

[43]. N.S. Aini, V.D. Kharisma, M.H. Widyananda, A.A. Ali Murtadlo, R.T. Probojati, D.D. Rahma Turista, M.B. Tamam, V. Jakhmola, D. Novaliendry, R.S. Mandeli, Bioactive compounds from purslane (Portulaca oleracea L.) and star anise (Illicium verum Hook) as SARS-CoV-2 antiviral agent via dual inhibitor mechanism: In silico approach, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[44]. N. Contreras, A. Alviz-Amador, I. Manzur-Villalobos, In silico study of dimethyltryptamine analogues against 5-HT1B receptor: Molecular docking, dynamic simulations and ADMET prediction, *Journal of Herbmed Pharmacology*, **2022**, *11*, 204-212. [Crossref], [Google Scholar], [Publisher]

[45]. R.T. Probojati, S.L. Utami, D.D.R. Turista, A. Wiguna, A. Wijayanti, Y. Rachmawati, A.F. Dibha,

A.A.A. Murtadlo, T. Hasan, P. Listiyani, B-cell epitope mapping of capsid L1 from human papillomavirus to development cervical cancer vaccine through in silico study, *SAINSTEK International Journal on Applied Science*, *Advanced Technology and Informatics*, **2022**, 1, 62-71. [Crossref], [Google Scholar]

[46]. A. Dibha, S. Wahyuningsih, A. Ansori, V. Kharisma, M. Widyananda, A. Parikesit, M. Sibero, R. Probojati, A. Murtadlo, J. Trinugroho, Utilization of secondary metabolites in algae Kappaphycus alvarezii as a breast cancer drug with a computational method, *Pharmacognosy Journal*, **2022**, *14*. [Crossref], [Google Scholar], [Publisher]

[47]. A.A.A. Murtadlo, P. Listiyani, S.L. Utami, S. Wahyuningsih, D.D.R. Turista, A. Wiguna, A. Wijayanti, Y. Rachmawati, A.F. Dibha, T. Hasan, Molecular docking study of nigella sativa bioactive compound as E6 inhibitor against human papillomavirus (HPV) infection, *SAINSTEK International Journal on Applied Science, Advanced Technology and Informatics*, **2022**, *1*, 32-38. [Crossref], [Google Scholar], [Publisher]

[48]. N. Bung, S.R. Krishnan, A. Roy, An in silico explainable multiparameter optimization approach for de novo drug design against proteins from the central nervous system, *Journal of Chemical Information and Modeling*, **2022**, *62*, 2685-2695. [Crossref], [Google Scholar], [Publisher]

[49]. S. Choudhary, Y.S. Malik, S. Tomar, Identification of SARS-CoV-2 cell entry inhibitors by drug repurposing using in silico structure-based virtual screening approach, *Frontiers in Immunology*, **2020**, *11*, 1664. [Crossref], [Google Scholar], [Publisher]

[50]. K. Anam, B. Prabowo, M.T. Kusuma, S. Winarsih, T.Y.M. Raras, S.R. Prawiro, Multi epitopes potential on surface sars-cov-2 protein as a covid-19 vaccine candidate, *Research Journal of Pharmacy and Technology*, **2022**, *15*, 1437-1442. [Crossref], [Google Scholar], [Publisher]

[51]. H. Guo, H. Zeng, C. Fu, J. Huang, J. Lu, Y. Hu, Y. Zhou, L. Luo, Y. Zhang, L. Zhang, Identification of sitogluside as a potential skin-pigmentationreducing agent through network pharmacology, *Oxidative Medicine and Cellular Longevity*, **2021**, *2021*. [Crossref], [Google Scholar], [Publisher]

[52]. V.D. Kharisma, A. Agatha, A.N.M. Ansori, M.H. Widyananda, W.C. Rizky, T.G.A. Dings, M. Derkho, I. Lykasova, Y. Antonius, I. Rosadi, Herbal combination from Moringa oleifera Lam. and Curcuma longa L. as SARS-CoV-2 antiviral via dual inhibitor pathway: A viroinformatics approach, *Journal of Pharmacy & Pharmacognosy Research*, **2021**, *10*, 138-146. [Crossref], [Google Scholar], [Publisher]

[53]. M.E. Ullah, S.W. Naw, A.A.A. Murtadlo, M.B. Tamam, R.T. Probojati, Molecular mechanism of black tea (Camellia sinensis) as SARS-CoV-2 spike glycoprotein inhibitor through computational approach, *SAINSTEK International Journal on Applied Science*, *Advanced Technology and Informatics*, **2022**, 1, 20-25. [Google Scholar] [54]. W. Yin, X.E. Zhou, D. Yang, P.W. de Waal, M. Wang, A. Dai, X. Cai, C.-Y. Huang, P. Liu, X. Wang, Crystal structure of the human 5-HT1B serotonin receptor bound to an inverse agonist, *Cell Discovery*, **2018**, *4*, 12. [Crossref], [Google Scholar], [Publisher]

[55]. M. Zweckstetter, A. Dityatev, E. Ponimaskin, Structure of serotonin receptors: molecular underpinning of receptor activation and modulation, *Signal Transduction and Targeted Therapy*, **2021**, *6*, 243. [Crossref], [Google Scholar], [Publisher]

[56]. P. Xu, S. Huang, H. Zhang, C. Mao, X.E. Zhou, X. Cheng, I.A. Simon, D.-D. Shen, H.-Y. Yen, C.V. Robinson, Structural insights into the lipid and ligand regulation of serotonin receptors, *Nature*, **2021**, *592*, 469-473. [Crossref], [Google Scholar], [Publisher]

[57]. F. Shojaie, Quantum computations of interactions of most reactive tricyclic antidepressant drug with carbon nanotube, serotonin and norepinephrine, *Chemical Methodologies*, **2020**, *4*, 447-466. [Crossref], [Google Scholar], [Publisher]

Copyright © 2024 by SPC (Sami Publishing Company) + is an open access article distributed under the Creative Commons Attribution License (CC BY) license (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.